



Assessment of Her2/Neu status by Silver Insitu Hybridization in Immunohistochemistry Equivocal Cases of Invasive Breast Cancer-Cross Sectional Study in a Sample of Iraqi Patient

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Original Article

Summary

Determination of the Her-2/neu gene amplification status is crucial in breast cancer because it allows selecting patients who would benefit from treatment where the treatment agent designed to recognize and bind to HER2 protein and according to published clinical studies. This study aimed to assess Her2/Neu status by Silver insitu hybridization in immunohistochemistry equivocal cases of invasive breast carcinoma. We conducted this cross-sectional during the period from January to August 2021. The study included 50 Iraqi women with proved diagnosed invasive breast cancer with equivocal IHC result for Her2\Neu. Scoring of Her2 gene amplification was classified according to American society of clinical oncology college of American pathologist(ASCOS CAP) 2018 guidline the obtained results where correlated with given data of patient age and hormone receptor(ER,PR) status. Results showed a mean age of patients of 47.2 \pm 9.9 (range: 30 – 75) years. Ten patients (20%) were ER-, PR- and (80%) were ER+, PR+. Among the 50 cases, (62%) HER2/neu was not amplified. On both univariate and binary regression analysis,HER2/neu status was significantly associated with ER-, PR- independent of age , (OR=5.13, P. value = 0.035). In conclusions, Silver insitu hybridization is useful in assessment of HER2/ neu status in immunohistochemistry equivocal cases of invasive breast carcinoma. ER negative ,PR negative status was significantly associated with amplification of HER2/neu independent of patients' age .

Keywords: Breast cancer, Hormonal Receptors, HER2/neu, Silver insitu hybridization, immunohistochemistry

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1. INTRODUCTION

Breast cancer is the most common cancer among women worldwide, both in developed and developing countries, accounting for 24% of new cancer cases and 15% of cancer deaths in women (3-5) In Iraq, the annual incidence of breast cancer has increased significantly with an average incidence rate of 37.9/100000 in 2019. The the age-adjusted incidence rate in Iraq found to be greater than that in other countries like Turkey, Iran, Saudi Arabia and Bahrain(6,7). There are many risk factors for breast cancer such as female gender, older age, late age of menopause, contraceptive hormonal methods, benign breast lesions, obesity, smoking and others (8). Screening, early detection and diagnosis of breast cancer play a crucial role in success of treatment of breast cancer (9,10). Earlier studies and literatures documented that Human epidermal growth factor receptor 2(Her 2) overexpressing breast cancer is known to be more aggressive disease and associated with poor prognosis. From other point of view, Her2/neu overexpression considered as predictor of response to endocrine chemotherapy (11,12). On the other hand, neoadjuvant chemotherapy for breast cancer have shown to result in alteration in HER2/neu status by immunohistochemistry, but they have stable status of gene amplification by fluorescence in situ hybridization (FISH). Determination of HER-2/neu oncogene amplification has become necessary for selection of breast cancer patients for trastuzumab (Herceptin) therapy. Fluorescence in situ hybridization (FISH) is currently regarded as a gold standard method for detecting HER-2/neu amplification, but it is not very practical for routine histopathological laboratories. We evaluated a new modification of in situ hybridization, the Silver in situ hybridization (SISH), which enables detection of HER-2/neu gene copies with conventional peroxidase reaction. Clinical importance of HER-2 diagnostics got more attention and awareness by clinician and oncologist with the increased new anti-cancer treatment, HER-2 assays are now considered a substantial an part of diagnostic methods of breast cancer paralleled to hormonal receptors(15). The earliest studies of HER-2 used Southern and Western blotting for detection of HER-2 gene amplification and protein overexpression. These methods are not well suited for routine diagnostics and have been replaced by immunohistochemistry (IHC) and fluorescence in situ hybridization (FISH). A vast majority of HER-2 studies has been done using IHC, which detects the HER-2 protein overexpression on the cell membrane. Without HER-2

oncogene amplification, the protein expression is low and undetectable by IHC. However, IHC is subject to a number of technical artifacts and sensitivity differences between different antibodies and tissue pretreatments. 3 Standardized reagent kits have recently been introduced (such as HercepTest), but mixed results have been reported from their methodological comparisons (16,17). FISH quantifies the number of gene copies in the cancer cell nucleus. Since initial applications to detect HER-2 amplification by FISH, a number of reports have verified its accuracy both in freshly frozen and paraffin-embedded tumor material. FISH is done either using single-color (HER-2 probe only, DAKO, Copenhagen, Denmark) or as a dual-color hybridization (using HER-2 and chromosome 17 centromere probes simultaneously), the latter making it easier to distinguish true HER-2 amplification from chromosomal aneuploidy. FISH from entire cells (cultured cells, pulverized tissue, or imprint touch specimens from tumors) is considered straightforward, but the use of tissue sections complicates the quantitative nature of FISH because of nuclear truncation (slicing) (18,19). The main difficulty for adopting FISH in clinical diagnostics is the need to use fluorescence microscopy, which is not done in most routine diagnostic laboratories. Evaluation of FISH requires a modern epifluorescence microscope equipped with high-quality $\times 60$ and $\times 100$ oil immersion objectives and multi-bandpass fluorescence filters. Moreover, because the fluorescence signals fade within a few weeks, the hybridization results must be recorded with expensive digital cameras(18,19).

Principles of HER2 testing (20–22).

There are a large number of methods available for the detection and measurement of HER2/neu expression; some are based on the overexpression of the protein, with the use of immunohistochemistry, and others evaluate the amplification of the gene with different techniques, such as: FISH, SISH and real-time PCR. There is also the possibility of using the ELISA (Enzyme Linked Immunoabsorvent Assay) technique to measure the antigen in the serum according to the NCCN guide (2016)

Immunohistochemistry (IHC). (23,24)

IHC is a semi-quantitative technique used for the quantification of protein expression; reveals different epitopes of the protein present on the cell surface, and is the most widely used technique to detect and quantify the HER2/neu protein in the first instance. This technique detects the HER2/neu receptor on the cell membrane using antibodies that bind

to the HER2/neu receptor. This receptor is the target to which the therapeutic agent trastuzumab binds, and therefore the overexpression of this protein should predict the response to this agent. There are many variables that affect the result of immunohistochemistry: fixation, tissue storage, antigen retrieval, type of antibody, measurement system, and interpretation variability among observers. Prolonged fixation in formalin causes changes in protein configuration that lead to masking of antigenic sites and can cause false negatives. The use of 10% buffered formalin with a fixation period of 6 to 12 hours is recommended. It should be taken into account that prolonged storage of paraffin-embedded tissues can cause false negatives, associated with antigen degradation The interpretation of the results is based on the assessment of the intensity of the staining of the cell membranes and the percentage of positive tumor cells. Results are reported on a scale of 0 to 3 + as follows:

IHC 0.1+: HER2 negative :incomplete membrane staining that is faint/barely perceptible in more than 10% of tumor cell

IHC 2+: equivocal result, weak to moderate complete membrane staining observed in more than 10% of tumor cell in this case FISH tests are performed or a new IHC or FISH test is requested

IHC 3+: HER2 positive: circumferential staining that is complete intense in more than 10% of tumor cell .

Fluorescent in situ hybridization (FISH). FISH is a molecular cytogenetic method that allows the number of copies of a gene to be quantified. FISH is currently considered the gold standard for evaluating HER2/neu amplification, it has a sensitivity and specificity of 98% and 100%, respectively. The currently recommended algorithm for HER2/neu evaluation is as follows:

Initial screening with immunohistochemical test.

All patients with a score of 2+ should be Reffered to FISH, due to the high rate of false positives. Patients with a score of 3 + by immunohistochemistry and those with a score of 2 + who have FISH amplification are considered eligible for treatment with trastuzumab.

Patients with a score of 0 or 1+ and those with a score of 2+ who do not amplify with FISH are considered ineligible (25,26)

Silver in situ hybridization (SISH)

Silver-enhanced in situ hybridization (SISH) has been developed as alternative method to FISH and CISH for HER2 determination .SISH is a novel bright-field in situ hybridization technique similar to CISH. It is a fully automated system developed by (Ventana Medical System) that improves the efficiency and consistency of brigh field in situ hybridization and reducing the risk of error.

Interpretation of results done as follow

Dual Probe ISH Group Definitions:

Group 1 = HER2/CEP17 ratio ≥ 2.0 ; ≥ 4.0 HER2 signals/cell

Group 2 = HER2/CEP17 ratio ≥2.0; <4.0 HER2 signals/cell

Group 3 = HER2/CEP17 ratio $<2.0; \ge 6.0$ HER2 signals/cell

Group 4 = HER2/CEP17 ratio <2.0; \geq 4.0 and <6.0 HER2 signals/cell

Group 5 = HER2/CEP17 ratio <2.0; <4.0 HER2 signals/cell

| Result | Criteria(dual-probe assay) | |
|----------|---|--|
| Negative | Group 5 | |
| Negative | Group 2 and concurrent IHC 0-1+ or 2+ Group 3 and concurrent IHC 0-1+ Group 4 and concurrent IHC 0-1+ or 2+ | |
| Positive | Group 2 and concurrent IHC 3+ Group 3 and concurrent IHC 2+ or 3+ Group 4 and concurrent IHC 3+ | |
| Positive | Group 1 | |

Reporting Results of HER2 Testing by In Situ Hybridization (dual-probe assay)

2. PATIENTS and METHODS

This was a cross-sectional study conducted during the period from january 2021 to August 2021. Included 50 Iraqi women with proved diagnosed invasive breast cancer with equivocal IHC result for Her2\Neu that where referred to ISH unit for study of gene amplification and the tumor tissues were tested for receptors expression and Her2/Neu

status; automated SISH for consecutive slides from the same paraffin blocks as for Her2/Neu IHC where stained according to manufactures protocol for INFORM Her2 Dual ISH DNA probe cocktail on ventona Benchmark R XT slide stainer.

Staining protocol for HER2 ISH by ventana BenchMark XT automated slide stainer

| Deparaffinization | selected |
|-----------------------------|--------------------------------|
| Extended deparaffinization | not selected |
| Cell conditioning | selected cell conditioning CC2 |
| | mild CC2 8minutes |
| | standard CC2 12 minutes |
| | extended CC2 8 minutes |
| ISH protease3 | 16 minutes |
| Denaturation | 20 minutes |
| Hybridization | 6 hours |
| Stringency wash temperature | 72Deg C |
| SISH multimer | 16 minutes |
| Silver chromogen | 4 minutes |
| Red ISH multimer | 24 minutes |
| Red chromogen | 8 minutes |
| Counterstain | Hematoxylin II-8 minutes |
| Post counterstain | Bluing reagent 4 mi |

Statistical analysis:

Data of the 50 patients were entered and managed using Microsoft Excel program version 2016. Analysis of data and extraction of findings and correlations was managed with the statistical package for social sciences (SPSS) software for windows, version 26. Descriptive statistics presented as frequency (No.) and percentages (%) for categories. Also mean and standard deviation was calculated for age of the patients as scale variable. Pie – chart distribution used to demonstrate the distribution of the studied group according to the HER2/neu status. Chi square test used to assess the significance of association between HER2/neu status from one side and each of ER, PR receptor status and age of patient from

the other side. Further analysis was performed using binary regression analysis to control the possible effect of age on the correlation between ER,PR receptors status and HER2/neu status . In all statistical analyses the level of significance of ≤ 0.05 considered significant. Finally, results expressed in tables ,figure and picture with interpretation for each using Microsoft Office Word Program version 2016.

3. RESULTS

A total of 50 patients were enrolled in this study with a mean age of 47.2 ± 9.9 (range: 30 – 75) years, moreover a significant higher proportion of patients (90%) were older than 40 years, (P. value < 0.001), (Table 1). Hormonal status of the studied group revealed that 10 patients (20%) were ER-, PR- and the remaining 40 patients (80%) were ER+, PR+, with significant higher frequent ER+, PR+ status, (P<0.001), (Table 2). Distribution of the studied group according to HER2/neu status using Pie-chart showed not amplified HER2/neu status in 31 (62%) of cases and amplified HER2/neu status in 19 (38%) of cases, indicated significant higher frequency of not amplified HER2/neu status, (P. value = 0.002). Relationship between ER,PR receptors status and HER2/neu status of the studied group is shown in (Table 3). where a significant association was found between ER-, PR- status and amplified HER2/neu status , (P. value = 0.020) on the other hand , calculation of odds ratio indicated that patients with ER-, PR- were about 5.44 folds more likely to have amplified HER2/neu , (Odds ratio = 5.44). Further comparison was performed to assess possible correlation between age and HER2 status , this comparison was statistically insignificant, (P. value > 0.05), (Table 4).

| ¥ | | | |
|-------------------|--------------|-------|----------|
| Age (year) | No. | % | P. value |
| \leq 40 | 10 | 20.0 | |
| 41 - 50 | 28 | 56.0 | < 0.001 |
| > 50 | 12 | 24.0 | |
| Total | 50 | 100.0 | |
| Mean age \pm SD | 47.2 ± 9.9 | | |
| Range | 30 - 75 | | |

Table1. Age distribution of the studied group

SD: standard deviation of mean

| ER,PR receptors status | No. | % | P. value | |
|------------------------|-----|-------|----------|--|
| ER-, PR- | 10 | 20.0 | < 0.001 | |
| ER+, PR+ | 40 | 80.0 | < 0.001 | |
| Total | 40 | 100.0 | | |

Table 2. Hormone receptors status of the studied group

Table 3. Cross-tabulation for the relationship between hormone receptor status and HER2/neu status of the studied group

| | HER2/neu status | | | | | |
|--|-----------------|------|---------------|------|-------|-------------|
| | Amplified | | Not amplified | | Total | P. value |
| ER,PR receptors | No. | % | No. | % | | |
| ER-, PR- | 7 | 70.0 | 3 | 30.0 | 10 | 0.020 |
| ER+, PR+ | 12 | 30.0 | 28 | 70.0 | 40 | 0.020 |
| Total | 7 | 14.0 | 3 | 6.0 | 10 | |
| Odds ratio for ER-, PR- state = 5.44 | | | | | | |

Table 4. Cross-tabulation for the relationship between age and HER2/neu status of the studied group

| Age (year) | HER2 status | | | | | D |
|------------|-------------|------|---------------|------|-------|-------------|
| | Amplified | | Not amplified | | Total | P. value |
| | No. | % | No. | % | | |
| \leq 40 | 4 | 40.0 | 6 | 60.0 | 10 | |
| 41 - 50 | 13 | 46.4 | 15 | 53.6 | 28 | 0.204 |
| > 50 | 2 | 16.7 | 10 | 83.3 | 12 | |
| Total | 19 | 38.0 | 31 | 62.0 | 50 | |

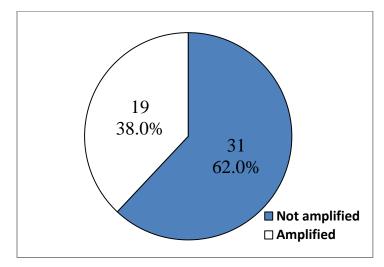


Figure 1. Pie-chart showing the HER2/neu status of the studied group (P. value = 0.002)

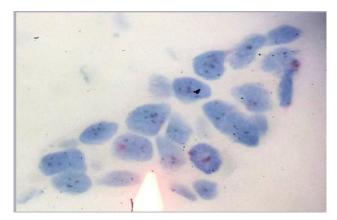


Figure 2. One-Two copies of chromosome 17 red ISH signal present(60x)

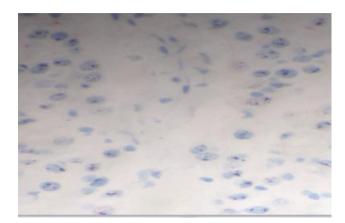


Figure 3. HER2 amplified with presence of HER2 clusters(40*x*)

4. DISCUSSION

The importance of determining HER2 status lies in the fact that those patients who have breast cancer in advanced stages of the disease and who also present Her2 amplification have greater resistance to conventional chemotherapy and hormonal treatment, in addition to a lower survival rate. The reason why expression of HER2/neu is amplified at 20 to 30 is unknown. This gene is associated with a poor prognosis and survival of the disease; however, currently the monoclonal antibody called Herceptin or trastuzumab inhibits the proliferation of cells in which this oncogene is overexpressed (30-33). Determination of the Her-2/neu gene amplification status is crucial in breast cancer because it allows selecting patients who would benefit from treatment where the treatment agent designed to recognize and bind to HER2 protein and according to published clinical studies, it reduces the risk of early-stage cancer recurrence and reduces the relative risk of death by 20% in patients with metastatic breast cancer (34,35). There are different ways to detect the amplification of the Her-2/neu gene or the increase in the expression of the HER2 protein in the tumor. If you want to detect gene amplification, you can use Southern Blot, fluorescent in situ hybridization (FISH) or PCR; for transcript enhancement Northern Blot, and for HER2 protein enhancement immunohistochemistry (IHC). The technique chosen by Pathology laboratories for being sustainable cost/benefit is IHC. The limitations of this method are reflected in a group of tumors with an undefined 2+ result (intermediate results between positive 3+ and negative 0/1+). In these cases, FISH is considered a gold standard (24,25,27,29).

The current study aimed to assess Her2/Neu status by Silver insitu hybridization in immunohistochemistry equivocal cases of invasive breast carcinoma, among sample of 50 Iraqi breast cancer patient. The age distribution of the studied group revealed that majority of the patients were older than 40 years, which indicated that the risk of breast cancer is higher in older age women. These finding was not unexpected as the older age is important risk factor of breast cancer where the risk increases significantly with advancing age. Our findings consistent with epidemiological studies regarding the risk factors of breast cancer aged between 50-59 years and the risk increases with the older age (36). According to the American cancer Society , only 4% of diagnosed women with breast cancer are younger

than 40 years (37). In the present study both ER and PR were negative in 20% of the cases while both positive in majority (80%) of cases. These findings agreed that reported in previous studies; Hu et al documented that 80% of breast cancers were hormonal receptor positive (38). On the other hand, heterogeneous, i.e. ER-/PR+, expression of receptor is not common subtype, and PR expression have shown to be not associated with prognosis of ER-breast cancer (39). However, assessment of hormone receptors expression is one of the crucial components of pathological assessment of breast cancer (40). From other point of view, there is a strong evidence about the importance of the biologic, prognostic and predictive role of ER expression in breast cancer, but, the additive role of PR assessment is still controversial (41). Nonetheless, the American Society of Clinical Oncology and American Pathologist College recommended the testing of both ER and PR(39). Regarding HER2/neu status in our study we found that it was not amplified in 32% and amplified in the remaining 38%. Earlier study conducted by Panvichian et al. detected HER2 expression with IHC on formalin fixed paraffin embedded sections of 37 breast carcinoma (42).

Another study conducted by Murthy et al. found that 73.5% of the IHC 2+ cases were negative for HER2/new amplification, 25% were positive and only 2.2% were equivocal. Thus they concluded that IHC HER2 equivocal patients were heterogenous group and needed FISH for more precise assessment . The heterogeneity between IHC and FISH could be attributed to the polysomy 17 and HER2/neu genetic heterogeneity in equivocal cases (43). Furthermore, Panjwani et al. confirmed that IHC is a wise 1st step to screen tissue samples HER2/neu status to determine about the demand for FISH test as gold standard for assessing HER2/neu status in IHC equivocal breast cancer patients(44).

Also we found that patients with ER-/PR- expression were about 5.4 fold more likely to have amplified HER2/neu , (Odds ratio = 5.44). Moreover, we analyzed the association between ER, PR receptors expression and HER2/neu status after adjustment for age using regression analysis and found that HER2/neu status still significantly associated with ER-, PR-independent of age , (OR=5.13, P. value = 0.035). Our findings consistent with that reported by Huang et al. (45) who studied 1362 breast cancer cases and found that ER, PR were independent. We found no significant association between HER2/neu status and age of the patients, however, previous studies did not found significant association with the age but

Huang et al. found significant inverse association between HER2/neu and age in patients older than 45 years but not in younger group.

5. CONCLUSIONS

As it well documented in previous literature we concluded that frequency of breast cancer increases with advancing age. There is higher proportion of ER+, PR+ status among the studied group. ER, PR and HER2/neu status were significantly correlated and the ER negative ,PR negative status was significantly associated with amplification of HER2/neu independent of patients' age. Silver insitu hybridization is useful in assessment of HER2/ neu status in immunohistochemistry equivocal cases of invasive breast carcinoma. However, we recommended further studies with larger sample size to get further assessment.

REFERENCES

- 1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209–49.
- 2. Wu C, Li M, Meng H, Liu Y, Niu W, Zhou Y, et al. Analysis of status and countermeasures of cancer incidence and mortality in China. Sci China Life Sci. 2019;62(5):640–7.
- 3. Heer E, Harper A, Escandor N, Sung H, McCormack V, Fidler-Benaoudia MM. Global burden and trends in premenopausal and postmenopausal breast cancer: a population-based study. Lancet Glob Heal. 2020;8(8):e1027–37.
- 4. Lei S, Zheng R, Zhang S, Chen R, Wang S, Sun K, et al. Breast cancer incidence and mortality in women in China: temporal trends and projections to 2030. Cancer Biol Med. 2021;18(3):900.
- 5. Lei S, Zheng R, Zhang S, Wang S, Chen R, Sun K, et al. Global patterns of breast cancer incidence and mortality: A population-based cancer registry data analysis from 2000 to 2020. Cancer Commun. 2021;41(11):1183–94.
- 6. Hashim HT, Ramadhan MA, Theban KM, Bchara J, El-Abed-El-Rassoul A, Shah J. Assessment of breast cancer risk among Iraqi women in 2019. BMC Womens Health. 2021 Dec 15;21(1):412.
- 7. Al-Hashimi MMY. Trends in Breast Cancer Incidence in Iraq During the Period 2000-2019. Asian Pacific J cancer Prev APJCP. 2021;22(12):3889–96.
- 8. Momenimovahed Z, Salehiniya H. Epidemiological characteristics of and risk factors for breast cancer in the world. Breast Cancer Targets Ther. 2019;11:151.
- 9. Naeem M, Hayat M, Qamar SA, Mehmood T, Munir A, Ahmad G, et al. Risk factors, genetic mutations and prevention of breast cancer. Int J Biosci. 2019;14(4):492–6.
- 10. Ginsburg O, Yip C, Brooks A, Cabanes A, Caleffi M, Dunstan Yataco JA, et al. Breast cancer early detection: A phased approach to implementation. Cancer. 2020;126:2379–93.
- 11. Ferretti G, Felici A, Papaldo P, Fabi A, Cognetti F. HER2/neu role in breast cancer: from a prognostic foe to a predictive friend. Curr Opin Obstet Gynecol. 2007;19(1):56–62.
- 12. Li P, Liu T, Wang Y, Shao S, Zhang W, Lv Y, et al. Influence of Neoadjuvant Chemotherapy on

HER2/neu Status in Invasive Breast Cancer. Clin Breast Cancer. 2013;13(1):53-60.

- 13. Sasanpour P, Sandoughdaran S, Mosavi-Jarrahi A, Malekzadeh M. Predictors of pathological complete response to neoadjuvant chemotherapy in Iranian breast cancer patients. Asian Pacific J cancer Prev APJCP. 2018;19(9):2423.
- 14. Ross JS, Fletcher JA, Linette GP, Stec J, Clark E, Ayers M, et al. The Her-2/neu gene and protein in breast cancer 2003: biomarker and target of therapy. Oncologist. 2003;8(4):307–25.
- 15. Furrer D, Paquet C, Jacob S, Diorio C. The Human Epidermal Growth Factor Receptor 2 (HER2) as a prognostic and predictive biomarker: Molecular insights into HER2 activation and diagnostic implications. Cancer Progn. 2018;
- 16. Jimenez RE, Wallis T, Tabasczka P, Visscher DW. Determination of Her-2/Neu status in breast carcinoma: comparative analysis of immunohistochemistry and fluorescent in situ hybridization. Mod Pathol. 2000;13(1):37–45.
- 17. Mitchell MS, Press MF. The role of immunohistochemistry and fluorescence in situ hybridization for HER2/neu in assessing the prognosis of breast cancer. In: Seminars in oncology. 1999. p. 108–16.
- 18. Huber D, von Voithenberg LV, Kaigala G V. Fluorescence in situ hybridization (FISH): history, limitations and what to expect from micro-scale FISH? Micro Nano Eng. 2018;1:15–24.
- 19. Jiang J. Fluorescence in situ hybridization in plants: recent developments and future applications. Chromosom Res. 2019;27(3):153–65.
- 20. Rakha EA, Pinder SE, Bartlett JMS, Ibrahim M, Starczynski J, Carder PJ, et al. Updated UK Recommendations for HER2 assessment in breast cancer. J Clin Pathol. 2015;68(2):93–9.
- 21. Lin L, Sirohi D, Coleman JF, Gulbahce HE. American society of clinical oncology/college of American pathologists 2018 focused update of breast cancer HER2 FISH testing GuidelinesResults from a national reference laboratory. Am J Clin Pathol. 2019;152(4):479–85.
- 22. Wolff AC, Hammond MEH, Allison KH, Harvey BE, Mangu PB, Bartlett JMS, et al. Human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline focused update. Arch Pathol Lab Med. 2018;142(11):1364–82.
- 23. Duraiyan J, Govindarajan R, Kaliyappan K, Palanisamy M. Applications of immunohistochemistry. J Pharm Bioallied Sci. 2012;4(Suppl 2):S307.
- 24. Buchwalow IB, Böcker W. Immunohistochemistry. Basics and Methods. 2010;1:1–149.
- 25. Levsky JM, Singer RH. Fluorescence in situ hybridization: past, present and future. J Cell Sci. 2003;116(14):2833–8.
- 26. Chrzanowska NM, Kowalewski J, Lewandowska MA. Use of fluorescence in situ hybridization (FISH) in diagnosis and tailored therapies in solid tumors. Molecules. 2020;25(8):1864.
- 27. Zhao J, Wu R, Au A, Marquez A, Yu Y, Shi Z. Determination of HER2 gene amplification by chromogenic in situ hybridization (CISH) in archival breast carcinoma. Mod Pathol. 2002;15(6):657–65.
- 28. Bartlett JMS, Campbell FM, Ibrahim M, Wencyk P, Ellis I, Kay E, et al. Chromogenic in situ hybridization: a multicenter study comparing silver in situ hybridization with FISH. Am J Clin Pathol. 2009;132(4):514–20.
- 29. Hanna WM, Kwok K. Chromogenic in-situ hybridization: a viable alternative to fluorescence in-

situ hybridization in the HER2 testing algorithm. Mod Pathol. 2006;19(4):481–7.

- 30. Dowsett M, Cooke T, Ellis I, Gullick WJ, Gusterson B, Mallon E, et al. Assessment of HER2 status in breast cancer: why, when and how? Eur J Cancer. 2000;36(2):170–6.
- 31. Carney WP. HER2 status is an important biomarker in guiding personalized HER2 therapy. 2005;
- 32. Bartlett J, Mallon E, Cooke T. The clinical evaluation of HER-2 status: which test to use? J Pathol A J Pathol Soc Gt Britain Irel. 2003;199(4):411–7.
- 33. Chivukula M, Bhargava R, Brufsky A, Surti U, Dabbs DJ. Clinical importance of HER2 immunohistologic heterogeneous expression in core-needle biopsies vs resection specimens for equivocal (immunohistochemical score 2+) cases. Mod Pathol. 2008;21(4):363–8.
- 34. Shah S, Chen B. Testing for HER2 in breast cancer: a continuing evolution. Patholog Res Int. 2011;2011.
- 35. Gutierrez C, Schiff R. HER2: biology, detection, and clinical implications. Arch Pathol Lab Med. 2011;135(1):55–62.
- 36. Kamińska M, Ciszewski T, Łopacka-Szatan K, Miotła P, Starosławska E. Breast cancer risk factors. Prz menopauzalny = Menopause Rev. 2015/09/30. 2015 Sep;14(3):196–202.
- 37. Society AC. Breast cancer facts & figures 2019-2020. Am Cancer Soc. 2019;1–44.
- 38. Hu T, Chen Y, Liu Y, Zhang D, Pan J, Long M. Classification of PR-positive and PR-negative subtypes in ER-positive and HER2-negative breast cancers based on pathway scores. BMC Med Res Methodol. 2021;21(1):108.
- 39. Hammond M, Hayes DF, Dowsett M, Allred DC, Hagerty KL. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer (unabridged version). Arch Pathol Lab Med. 2010;134(7):e48–72.
- 40. Hefti MM, Hu R, Knoblauch NW, Collins LC, Haibe-Kains B, Tamimi RM, et al. Estrogen receptor negative/progesterone receptor positive breast cancer is not a reproducible subtype. Breast Cancer Res. 2013;15(4):1–13.
- 41. Fuqua SAW, Cui Y, Lee A V, Osborne CK, Horwitz KB. Insights into the role of progesterone receptors in breast cancer. J Clin Oncol. 2015;23(4):931–2.
- 42. Panvichian R, Tantiwetrueangdet A, Wongwaisayawan S, Nampoon A, Lertsithichai P, Leelaudomlipi S. HER2 expression in breast cancer with nonamplified HER2 and gains of chromosome 17 centromere. Appl Immunohistochem Mol Morphol AIMM. 2012 Jul;20(4):367–74.
- 43. Murthy SS, Sandhya DG, Ahmed F, Goud KI, Dayal M, Suseela K, et al. Assessment of HER2/Neu status by fluorescence in situ hybridization in immunohistochemistry-equivocal cases of invasive ductal carcinoma and aberrant signal patterns: a study at a tertiary cancer center. Indian J Pathol Microbiol. 2011;54(3):532.
- 44. Panjwani P, Epari S, Karpate A, Shirsat H, Rajsekharan P, Basak R, et al. Assessment of HER-2/neu status in breast cancer using fluorescence in situ hybridization & immunohistochemistry: Experience of a tertiary cancer referral centre in India. Indian J Med Res. 2018;132(2):287–94.
- 45. Huang HJ, Neven P, Drijkoningen M, Paridaens R, Wildiers H, Van Limbergen E, et al. Association between tumour characteristics and HER-2/neu by immunohistochemistry in 1362

women with primary operable breast cancer. J Clin Pathol. 2005;58(6):611-6.

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